

B. Significance

B.1 Drug administration: a critical tool in biomedical research and clinical care

A critical capability in the selection of drugs, toxins, neurostimulators, and other biochemicals. Reliable chronic administration tools for mice are urgently needed given the wide diversity of drug-induced, transgenic, and knockout mouse models of human disease. In 2008, 25 million rats and mice were used in animal research in the US constituting 87% of all laboratory animals, the majority of which were mice.¹ Many novel drugs, such as biologics, siRNA, peptides, and small molecules, require special dosing regimens.² Current administration technologies are inefficient, resulting in wastefully high animal and personnel to animal charges. Only one in 10,000 drug candidates successfully obtain FDA approval.³ Rapid evaluation of efficacy, safety and toxicity in mice is now possible with our fully automated, remotely controlled drug dosing system.

B.2 Free and imperfect administration technologies for research animals

There is a long-standing animal need for cost-effective and advanced administration technologies for small animals capable of automated chronic dosing over a wide range of dosing regimens, especially in large scale studies requiring improved workflow, increased productivity, and automation. Manual administration (oral, intravenous, intraperitoneal) requires repeated handling and is most suitable for short-term dosing.⁴ The associated reduction of stress ("white coat" effects)⁵ may result in undesirable changes in physiological parameters (concentrations of corticosterone, glucose, growth hormone, or prolactin, heart rate, blood pressure, and behavior).⁶⁻¹¹ While chronic dosing is possible using a remote drug infusion pump and catheter either attached to the body as an implanted port, the latter poses risks of infection and entanglement.¹² Associated dosing responses include decreased respiratory activity, increased diuresis, aggression and mortality, and altered stress hormone response.¹³⁻¹⁵ The variability of administration to stress and environmental factors is well documented, including individual settings and ambulatory pumps.



Figure 1. Comparison of manual and automated drug administration. Left: Manual injection of drug into mouse. Right: Automated drug administration using the free-flow system.

Free-flowable pumps are available for small animals (Fig. 1). The **FlowFree** provides constant, single flow rate delivery to a compartment of the cage.¹⁶ Drug volume is limited (1-7 weeks) and flow rate cannot be changed. The **FlowFree** provides some electronic control of drug delivery (limited programmed steps, flow rates, and infusion locations) and is reliable. However, the pump is too large for mice, is always ON even when drug is not needed, and has only 6-month battery life. Pipelined and battery limitations necessitate risky surgical replacement of pumps for chronic dosing. Most importantly, the study cannot be synchronized to group housing, ambulatory dosing, and batch testing.

B.3 Outcomes improved in animal studies using telemetry

Indicatively used for monitoring physiological parameters in laboratory animals,¹⁷ reduces stress related to acquisition of measurements,¹⁸⁻²⁰ and eliminates restraint.²¹ Telemetry allows reduction of use of animals²²⁻²⁴ and 24-hour data collection without artificial environmental structures.²⁵ Telemetry benefits in improvement of study outcomes and animal welfare are paired with wireless implantable infusion systems. Automated wireless evaluation of multiple animals allows for higher throughput compared to manual administration.

C. Innovation

The **FlowFree**™ free-flow drug delivery system is the first and only wireless and tether-free administration system for mice (Fig. 2). Our system is a smart chamber where a network of implanted micro-pumps are remotely controlled from a single computing system. End users are animal researchers in academia and industry who need to control external experimental variables and labor costs, increase infection rates, manage drug toxicity with accurate dosing, reduce repeated implant surgeries, and select dosing on-demand to test in the right drug regimens over a controlled period.

The specific features that enable our innovative system are (1) low power **FlowFree** and high accuracy **FlowFree** electronics.

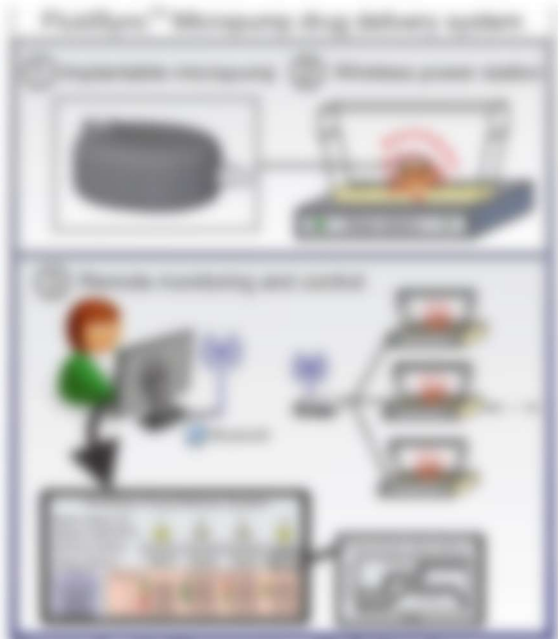


Figure 2. Schematic diagram of the FlowFree system. The system consists of a central computing system connected to a network of implanted micro-pumps in mice. The mice are housed in a smart chamber. The system is controlled by a single computing system, which is connected to a laptop and a monitor displaying data.